

EVALUATION OF CYTOTOXIC AND GENOTOXIC EFFECT OF THE TEXTILE DYE DIRECT BROWN ON *ALLIUM CEPA* L

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ABSTRACT

In the present study, the cytotoxic, genotoxic and mutagenic effects of the textile dye Direct Brown was evaluated using root tip cells of *Allium cepa*. Root length, mitotic indices and chromosomal aberrations were used as the test parameters. There was significant difference ($p < 0.05$) between mean root lengths of *A. cepa* exposed to different concentrations of the dye and the control. Root growth inhibition was concentration dependent and IC_{50} value was found as 200 ppm using regression plot. The mitotic index decreased significantly with dye treatments. Chromosomal aberrations such as sticky metaphase, disturbed metaphase, anaphasic bridge, disturbed anaphase, laggards and chromosome fragments were observed.

KEYWORDS: Chromosomal Aberrations, Cytotoxicity, Direct Brown, Genotoxicity, Textile Dyes

INTRODUCTION

Water pollution is a major threat which deepens the ecological crisis of the day. Excessive and indiscriminate use of synthetic textile dye stuffs has become increasingly a subject of environmental concern. These dyes can enter the environment through the industrial effluents of dye manufacturing plants. Moreover labors are getting exposed to these dyes during manufacturing and from textile dyeing and printing operations. Assessment of genotoxicity of dyes is therefore of utmost importance. Azo dyes, compounds characterized by the presence of one or more azo groups ($-N=N-$) – constitute the most important class in the textile industry [3], because they are one of the most easily synthesized dyes, showing both excellent fixation properties and permanence in fibers [24]. Azo dyes are known to be recalcitrant and highly toxic and also become harmful to the environment by the formation of aromatic amines (anilines), which are carcinogenic and/or mutagenic [14]. They are also used as reagents and biological stains in laboratories, are used in food industries and have more recent uses in laser, liquid crystal displays, ink-jet printers and electro-optical devices [16,4].

Plants can be considered as biosensors of genetic toxicity of the environmental pollutants. To the best of our knowledge, the diazo textile dye Direct Brown is not yet studied for its cytogenotoxic effects. Therefore, in the present investigation, the toxicity potential of the textile dye Direct Brown is studied by assessing their cytotoxicity, genotoxicity and mutagenicity on *Allium cepa*. Root length was used as the macroscopic parameter to assess the IC_{50} (inhibitory concentration) value and determine the cytotoxic effect of the dye on *A. cepa*. Microscopic parameters such as mitotic index and chromosomal aberrations were used to study the genotoxic and mutagenic effects induced by Direct Brown on *A. cepa*.

The Allium test is a sensitive test showing good correlation to other test systems. Thus positive results in the Allium test should be considered as a warning and also an indication that the tested chemical may be a risk to human health and to our ecosystem. Further, chromosome aberration studies are important for a better understanding of the action of chemicals on biological systems [5].

MATERIALS AND METHODS

Test Material

The textile diazo dye Direct Brown (DB) and healthy onion bulbs of almost same size and age were used for the study.

Macroscopic Studies for Cytotoxicity Evaluation of Direct Brown on *A. cepa*

Commercially available onion bulbs, *A. cepa* L. (2n = 16) were used as test system. A modified protocol of Fiskesjo[7] was used for *A. cepa* test setup. The roots already present at the base of the onion bulbs were carefully shaved off to expose the fresh meristematic tissues. For root growth evaluation, the bulbs were exposed directly to 50 ppm, 100 ppm, 200 ppm and 400 ppm treatment solutions of the test chemical (DB) prepared with distilled water.

For each treatment group, treatment solutions and control were changed every 24 hours till 72 hours. After 72 hours exposure, the roots of the bulbs were removed with a forceps and their lengths measured in centimeter with a meter scale. Data was subjected to statistical analysis and values are expressed as mean \pm S.D. Root growth inhibition was measured as IC₅₀, which is defined as the inhibitory concentration required for root length reduction to 50% of the control. IC₅₀ value of Direct Brown on *A. cepa* was assessed using the regression plot constructed with the help of Microsoft Excel computer programme. The significant difference (at P < 0.05) in the mean root lengths, was determined using analysis of variance (ANOVA) coupled with Q test [9].

Evaluation of Genotoxic and Mutagenic Effect of Direct Brown

Healthy bulbs of *A. cepa* were set for germination in sandy soil at 25°C to obtain roots. The root tips (1-1.5 cm length) from the germinated bulbs were treated with DB at different concentrations ranging from 50, 100, 200 and 400 ppm for 6 hours. The root tips treated in distilled water was used as control. After treatment, the root tips of control and experimental samples were thoroughly washed in distilled water and fixed in freshly prepared acetic acid: alcohol (1:3) solution. Chromosome preparations were made following haematoxylin squash technique [22]. Minimum of 5,000 cells from 5 root tips from 5 bulbs were analyzed to score the frequency of mitotic index (MI) and chromosomal aberrations (CA). The results are expressed as mean \pm SD. The statistical significance between control and experimental data were analyzed using ANOVA Tukey HSD.

RESULTS AND DISCUSSIONS

Cytotoxicity Evaluation of Direct Brown on *A. cepa*

A. cepa offers a good experimental model for *in vivo* evaluation of cytotoxicity and genotoxicity of chemical compounds and complex mixtures. Root length was considered as the macroscopic parameter for testing cytotoxicity (root growth inhibition) of the textile dye Direct Brown on *A. cepa* and the results of the study are presented in Table 1 and Figure 1. It was observed that the root growth was inhibited in a dose dependent manner.

Table 1: Effect of the Textile Dye Direct Brown on *A. cepa* Root Length

Concentration of the Dye (ppm)	Root Length (cm) after 72hrs
Control	8.26 \pm 0.29
50	6.52 \pm 0.31*
100	5.32 \pm 0.28*
200	3.84 \pm 0.30*
400	1.80 \pm 0.19*

Values are mean of five replicates and given as Mean \pm SD,

* significantly different from the control at P < 0.05 by one-way ANOVA coupled with Q-test

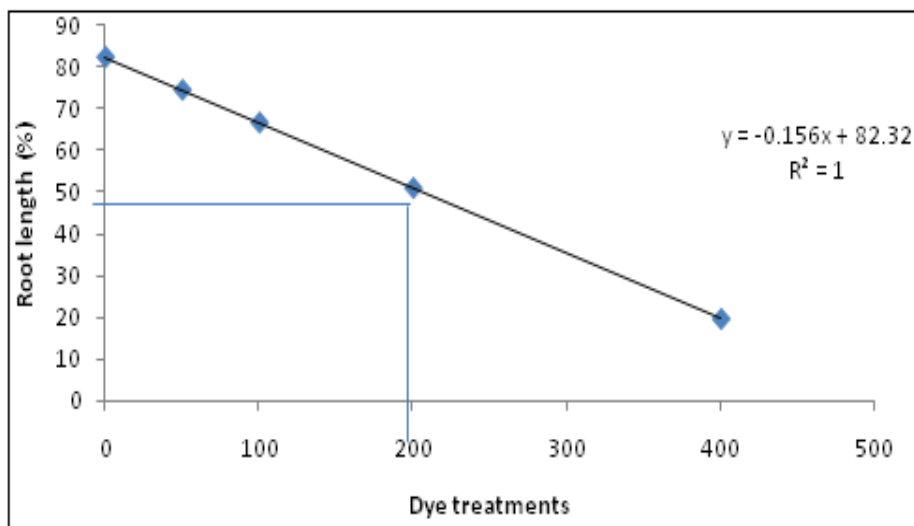


Figure 1: Assessment of IC₅₀ Value of Direct Brown Using Root Growth Inhibition Studies on *A. cepa*

The regression equation was determined and the IC₅₀ value assessed from a plot of root length as percentage of control against the treatment concentrations by using Microsoft Excel computer programme. The estimated IC₅₀ value of the dye was 200 ppm. Statistical analysis using analysis of variance (ANOVA) showed that there was a significant difference (at $P < 0.05$) in the mean root lengths of *A. cepa* exposed to different concentrations of the dye. Further analysis using Q test revealed that the root length of the control group *A. cepa* (treated with distilled water) was significantly different (at $P < 0.05$) from the root length of *A. cepa* exposed to all the different dye treatments.

Genotoxic and Mutagenic Effect of Direct Brown

Details of mitotic indices and frequency of chromosomal aberrations in *A. cepa* root tip cells after six hours of dye exposure are depicted in Table 2. Statistical analysis using ANOVA Tukey HSD comparison test revealed that all the dye treatments are significantly different from the control at $P < 0.05$. Significant reductions were observed in the mitotic index of the dye treated samples, while chromosomal aberration frequencies increased. The control exhibited the highest mitotic index (MI) of 9.29%, while cells exposed to 400 ppm dye concentration had the lowest MI (3.61%).

Chakraborty *et al.* suggested that a decreased MI of meristematic cells of *A. cepa* may be considered as a reliable method to determine the presence of cytotoxic compounds [2]. But Hoshina [10], opined that increased or decreased MI may serve as an important parameter for determination of the cytotoxicity level of a test compound. The present investigation and earlier observations indicate that the dyes and textile effluents exert effects on chromosomes and cell division [8, 19, 20, 21, 11, 17, 24]

Table 2: Mitotic Indices and Chromosomal Aberrations in *Allium cepa* Root Meristem Cells Treated with Direct Brown

Treatments	TAC	SD	AC	MN	Mitotic Index (Mean ± SD)	Chromosomal Aberration % (Mean ± SD)	Micronuclei (Mean ± SD)
Control	5375	502	15	4	9.29 ± 0.75	3.02 ± 1.50	0.76 ± 0.43
50 ppm	5350	290	45	7	5.37 ± 1.69 *	17.24 ± 8.66 *	1.28 ± 0.82
100 ppm	5275	311	56	8	5.86 ± 1.39 *	17.37 ± 3.31 *	1.55 ± 2.06
200 ppm	5135	314	53	16	5.99 ± 2.57 *	16.44 ± 3.20 *	3.21 ± 2.63
400 ppm	5300	195	51	5	3.61 ± 1.06 *	26.38 ± 5.34 *	1.01 ± 1.28

* - Significant at $P < 0.05$ (Control vs treated)

TAC – Total analyzed cells; SD – Showing division; AC – Aberrant cells; MN – Micronuclei

Besides reduction in mitotic index, genotoxicity analysis showed that different kinds of chromosomal aberrations were generated by the textile dye Direct Brown. Table 3 and 4 gives the indication of types, number and frequency of aberrations generated by Direct Brown. The number of total aberrant mitotic cells caused by all the concentrations of textile dye was apparently different from that of the control (Figure 2). The highest level of chromosomal aberration was observed in 400 ppm dye treatment (26.38 ± 5.34). 50, 100 and 200 ppm treatments had almost similar frequency and effect of chromosomal aberrations, while 400 ppm had an increased effect of chromosomal aberrations, since the total number of dividing cells were drastically less when compared with other treatments. No significant increase was observed in the frequencies of cells with micronuclei.

Total number of cells with alterations was significantly higher in dye treated samples than that of control, which is indicative of genotoxic nature of the dye. The most common chromosomal abnormalities in all treatments were sticky metaphase, disturbed metaphase, anaphasic bridge, disturbed anaphase and laggards (Figure 3).

Table 3: Frequency of Different Metaphasic Aberrations Observed in Dye Treated *Allium cepa* Root Cells

Dose (ppm)	AC	MA					
		CM		STM		DM	
		No	%	No	%	No	%
Control	15	1	6.67	3	20.00	4	26.67
50	45	3	6.67	6	13.33	12	26.67
100	56	2	3.57	25	44.64	8	14.29
200	53	1	1.89	11	20.75	5	9.43
400	51	2	3.92	15	29.41	7	13.73

AC – Aberrant cells; MA – Metaphasic aberrations; CM – C-metaphase; STM – Sticky metaphase; DM – Disturbed metaphase

Table 4: Frequency of Different Anaphasic Aberrations Observed in Dye Treated *Allium cepa* Root Cells

Dose (ppm)	AC	AA							
		LG		FG		AB		DA	
		No	%	No	%	No	%	No	%
Control	15	2	13.33	0	0	3	20.00	2	13.33
50	45	4	8.89	2	4.44	5	11.11	13	28.89
100	56	4	7.14	1	1.79	10	17.86	6	10.71
200	53	8	15.09	4	7.55	18	33.96	6	11.32
400	51	3	5.88	3	5.88	10	19.61	11	21.57

AC – Aberrant cells; AA – Anaphasic aberrations; LG – Laggards; FG – Fragments; AB – Anaphasic bridges; DA – Disturbed anaphase

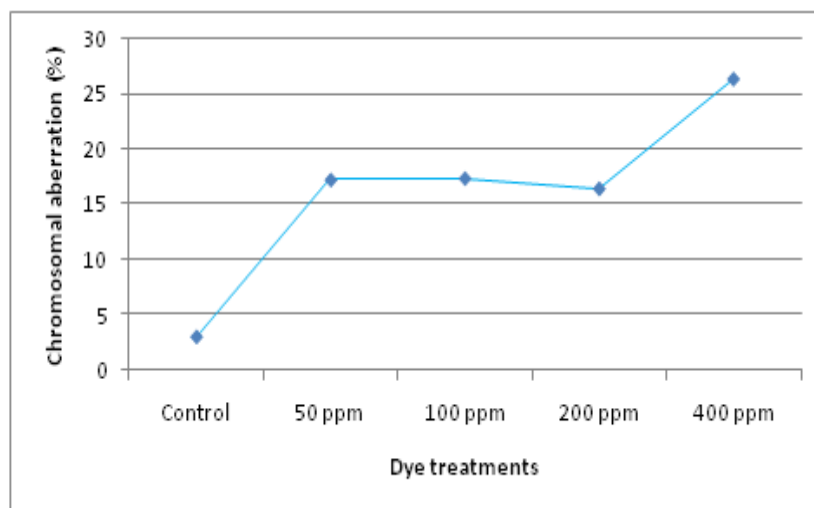


Figure 2: Effect of Dye Treatments on Chromosomal Aberrations

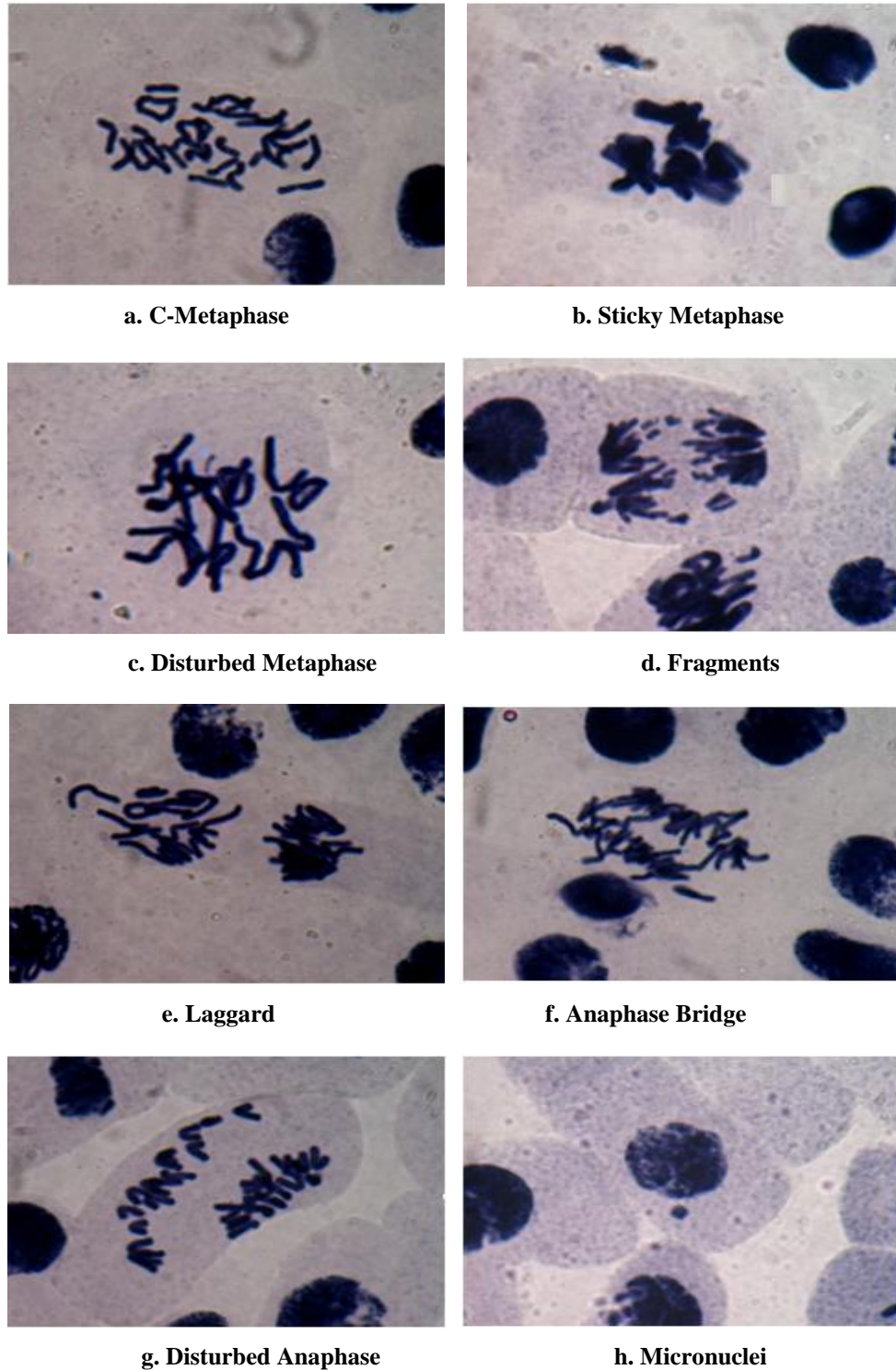


Figure 3: Genotoxic and Mutagenic Damages Observed in Meristematic Cells of *Allium cepa* Treated with Direct Brown

Determination of chromosomal aberrations can be considered as an important and efficient test for the investigation of genotoxicity potential of the textile dyes [1]. Sticky chromosomes indicate highly toxic, usually irreversible effect probably leading to cell death [5]. The presence of chromosome adherences (stickiness) may be a sign of genotoxic effect of the damage inducer, whose consequence of the action might lead to irreversible cell damage including cell death [13,23,12]. The present investigation also provides corroborative evidences to this. Highest frequency of sticky metaphase was observed in 100 ppm treatment of the dye (44.64% of the total aberrant cells). Highest frequency of anaphasic bridge was observed in 200 ppm dye treatments (34% of the total aberrant cells) along with highest frequency of

fragments (8%). Chromosome bridges and / or fragments result from chromosome- and chromatid- breaks. These aberrations may originate from either translocations or cohesive chromosome terminations [6].

The variation in the number of chromosomal aberration observed in this study was not dose dependent. This is not in accordance with Qian (2004), who reported that aberrant rate goes up with the concentrations [18]. But the results published by Odeigahet *al.* and Samuel *et al.* are supportive to the present investigation [15, 20]. According to Odeigahet *al.*, a possible explanation for this is that, with increasing concentration and consequently increasing toxicity, there is an inhibitory effect on cell division [15]. In the present study, even though the number of chromosomal aberrations were slightly less in the highest dose i.e., 400 ppm treatment when comparing with other treatments, it showed highest percentage of chromosomal aberrations and which is attributed to sharp decrease in mitotic index.

CONCLUSIONS

Significant reductions were observed in the mitotic index of the dye treated samples, while chromosomal aberration frequencies increased. The test compound induced different types of chromosomal aberrations on *A. cepa* such as sticky metaphase, disturbed metaphase, anaphasic bridge, disturbed anaphase, laggards and fragments. Considering all the types of chromosomal aberrations observed through the present study, it is concluded that the textile dye Direct Brown is cytotoxic, genotoxic and mutagenic in higher organisms. The present investigation thus emphasizes the need for the development of non-mutagenic dyes and efficient methods for the treatment of industrial effluents contaminated with synthetic dye stuffs in order to avoid the harmful effects on biological systems.

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REFERENCES

1. Carita, R. and Marin-Morales, M. A. 2008. Induction of chromosome aberrations in the *Allium cepa* test system caused by the exposure of seeds to industrial effluents contaminated with azo dyes. *Chemosphere*.**72**:722-725.
2. Chakraborty, R., Mukherjee, A. K. and Mukherjee, A. 2009. Evaluation of genotoxicity of coal fly ash in *Allium cepa* root cells by combining comet assay with the Allium test. *Environ. Monit. Assess.* **153**:351-357.
3. Chung, K. T. and Stevens, J. R. 1993. Degradation of azo dyes by experimental microorganisms and helminthes. *Environ. Toxicol. Chem.***12**:2121-2132.
4. Dapson, R. W. 2009. Benzidine-based dyes: effects of industrial practices, regulations and world trade on the biological stains market. *Biotechnic.Histochem.***84**:95-100.
5. Fiskesjo, G. 1985. The Allium test as a standard in environmental monitoring. *Hereditas.***102**:99-112.
6. Fiskesjo, G. 1993. Technical methods section. Allium test 1: A2-3 Day plant test for toxicity assessment by measuring the mean root growth of onions (*Allium cepa* L.). *Environ. Toxicol. Water Qual.***8**:461-470.
7. Fiskesjo, G. 1997. Allium test for screening chemicals; Evaluation of cytological parameters. In: Wang, W., Gorsuch, J. W. and Hughes, J. S. Plants for environmental studies. Lewis, New York. USA. pp:308-333.
8. Giri, A. K., Banerjee, T. S., Talukder, G. and Sharma, A. 1986. Effects of dyes (Indigo Carmine, Metanil Yellow, Fast Green FCF) and Nitrite *in vivo* on bone marrow chromosomes of mice. *Cancer Lett.***30**:315-320.

9. Gurumani, N. 2005. An introduction to biostatistics. Second revised edition. MJP Publishers. Chennai. pp 347-368.
10. Hoshina, M. M. 2002. Evaluation of a possible contamination of the waters of the Claro river municipality of Rio Claro, part of the Corumbatai river basin, with the mutagenicity tests using *Allium cepa*. 52f. Monograph (Bachelors and teaching degrees). State University of Sao Paulo, Rio Claro, SP.
11. Jadhav, S. B., Phugare, S. S., Patil, P. S. and Jadhav, J. P. 2011. Biochemical degradation pathway of textile dye remazol red and subsequent toxicological evaluation by cytotoxicity, genotoxicity and oxidative stress studies. *Int. Biodeteriorat. Biodegradat.* **65**:733-743.
12. Leme, D. M., Angelis, D. F. and Marin-Morales, M. A. 2008. Action mechanisms of petroleum hydrocarbons present in waters impacted by an oil spill on the genetic material of *Allium cepa* root cells. *Aquatic Toxicol.* **88**:214-219.
13. Marcano, L. and Del-Campo, A. 1995. Ultrastructural study of the nucleolus in *Allium cepa* L. onion meristematic populations treated on metabolic inhibitors. *Sci.* **3**:73-82.
14. Martins, M. A. M., Ferreira, I. C., Santos, I. M., Queiroz, M. J. and Lima, N. 2001. Biodegradation of bio-accessible textile azo dyes by *Phanerochaete chrysosporium*. *J. Biotechnol.* **89**:91-98.
15. Odeigah, P. G. C., Nurudeen, O. and Amund, O. O. 1997. Genotoxicity of oil field waste water in Nigeria. *Hereditas.* **126**:161-167.
16. Oomen, A. G., Versantvoort, C. H. M., Duits, M. R., van de Kamp, E. and van Twillert, K. 2004. Application of *in vitro* digestion models to assess release of lead and phthalate from toy matrices and azo dyes from textile. RIVM report 320102003/2004. Netherlands.
17. Oriaku, B. A., Otubanjo, O. A., Aderemi, A. O. and Otitolaju, A. A. 2011. Genotoxic end points in *Allium cepa* and *Clarias gariepinus* exposed to textile effluent. *Int. J. Environ. Protect.* **1**:48-52.
18. Qian, X. W. 2004. Mutagenic effects of chromium trioxide on root tips of *Vicia faba*. *J. Zhejiang. Univ. Sci.* **5**:1570-1576.
19. Roychoudhury, A. and Giri, A. K. 1989. Effects of certain food dyes on chromosomes of *Allium cepa*. *Mutat. Res.* **223**:313-319.
20. Samuel, B. O., Osuala, F. I. and Odeigah, P. G. C. 2010. Cytogenotoxicity evaluation of two industrial effluents using *Allium cepa* assay. *Afr. J. Environ. Sci. Technol.* **4**:21-27.
21. Sellappa, S., Prathyumnann, S., Joseph, S., Keyan, K. S. and Balachandar, V. 2010. Genotoxic effects of textile printing dye exposed workers in India detected by micronucleus assay. *Asian Pacific J. Cancer Preven.* **11**:919-922.
22. Sharma, A. K. and Sharma, A. 1980. Chromosome techniques. Theory and practice. Butterworths, London. pp.711.
23. Turkoglu, S. 2007. Genotoxicity of five food preservatives tested on root tips of *Allium cepa* L. *Mutat. Res.* **626**:4-14.
24. Ventura-Camarago, B. C., Maltempi, P. P. P. and Marin-Morales, M. A. 2011. The use of the cytogenetic to identify mechanisms of action of an azo dye in *Allium cepa* meristematic cells. *J. Environ. Anal. Toxicol.* **1**:109.

